





DetectX[®]

150913



MULTI-FORMAT KIT FOR

MULTIPLE SAMPLE TYPES

Serum Retinol Binding Protein Official Supplier Enzyme Immunoassay Kit

Catalog Number K004-H1

Species Independent

Sample Types Validated:

Serum, EDTA and Heparin Plasma

Please read this insert completely prior to using the product.

FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.

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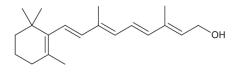
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WEB INSERT BACKGROUND



Retinol binding protein (RBP) is from a family of structurally related proteins that bind small hydrophobic molecules such as bile pigments, steroids, odorants, etc¹. RBP is a 21 kDa highly conserved, single-chain glycoprotein, consisting of 182 amino acids with 3 disulfide bonds, that has a hydrophobic pocket which binds retinol (vitamin A). The structure of retinol is shown below.



RBP binds retinol in a 1:1 stoichiometry, which serves to not only solubilize retinol but also protect it from oxidation. When in serum, the majority of RBP bound with retinol is reversibly complexed with transthyretin (prealbumin)². This complex then transports retinol to specific receptors of various tissues in the body. Vitamin A status is reflected by serum concentration as it is hemostatically controlled and does not fall until stores are dramatically reduced³⁻⁴.

RBP has been shown to be a useful surrogate marker for retinol because of the approximate 1:1 (molar) correlation between retinol and RBP in serum^{1, 5-6}, which implies that RBP may be used to assess and monitor vitamin A deficiency (VAD) in populations. The World Health Organization has estimated that 250 million children have moderate to severe VAD⁷ due to lack of adequate nutrition, and the rising cost of food staples around the world further exacerbates this problem. In addition to nutritional deficiencies, infectious stresses have been shown to depress retinol concentrations. Therefore, individuals with diseases such as cystic fibrosis⁸ and HIV-1⁹ also run the risk of VAD due to the infectious stresses that contribute to the disease.

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- 3. Goodman DS, Blaner WS. in The Retinoids, eds. Sporn MB, Roberts AB, Goodman DS, (Orlando: Aca demic Press, 1984) vol.2, 1-39.
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- Almekinder J, Manda W, Soko D, Lan Y, Hoover DR, Semba RD. "Evaluation of plasma retinol-binding protein as a surrogate measure for plasma retinol concentrations." Scand J Clin Lab Invest. 2000 May;60(3):199-203.
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- 7. World Health Organization, Division of Nutrition, "Global Prevalence of Vitamin A Deficiency," Micro nutrient Deficiency Information System, No.2 (1995) Geneva: WHO/UNICEF.
- 8. Duggan C, Colin AA, Agil A, Higgins L, Rifai N. "Vitamin A status in acute exacerbations of cystic fibrosis." Am J Clin Nutr. 1996 Oct;64(4):635-9.
- Baeten JM, Richardson BA, Bankson DD, Wener MH, Kreiss JK, Lavreys L, Mandaliya K, Bwayo JJ, McClelland RS. "Use of serum retinol-binding protein for prediction of vitamin A deficiency: effects of HIV-1 infection, protein malnutrition, and the acute phase response." Am J Clin Nutr. 2004;79:218-25.



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WEB INSERT Assay Principle



The DetectX[®] Retinol Binding Protein (RBP) Multi-Format Immunoassay kit is designed to quantitatively measure RBP present in serum and plasma samples. Please read the complete kit insert before performing this assay. This kit measures all isoforms of RBP.

The kit offers 2 standard curve ranges.

- For serum and plasma samples with normal RBP levels we recommend using 10 μL of standards or samples. The assay concentration range for RBP will be from 5,000 ng/mL to 39.1 ng/mL.
- For samples where RBP may be very low we recommend alternatively using 25 μL of standards or samples for an assay range of 2,500 ng/mL to 19.5 ng/mL

Standards or diluted samples are pipetted into a clear microtiter plate coated with an antibody to capture rabbit antibodies. RBP-peroxidase conjugate is added to the standards and samples in the wells. The binding reaction is initiated by the addition of the RBP polyclonal antibody to each well. After an hour incubation the plate is washed and substrate is added. The substrate reacts with the bound RBP-peroxidase conjugate. After a 30 minute incubation, the reaction is stopped and the intensity of the generated color is detected in a microtiter plate reader capable of measuring 450nm wavelength. The concentration of the RBP in the sample is calculated, after making a suitable correction for the dilution of the sample, using software available with most plate readers.

Please Note:

This kit has been reformatted and reformulated. Any changes to the format will be noted with this logo:

New Format

Watch Our EIA Video at:



RELATED **P**RODUCTS

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KITS Hemoglobin Detection Kit (2 Plate)

Catalog Number K013-H1



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WEB INSERT Supplied Components

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Coated Clear 96 Well Plate A clear plastic microplate with break-apart strips	1 Each coated with goat	Catalog Number X016-1EA anti-rabbit IgG.
RBP Standard A stock solution of native human RBP at 100 µg	60 μL g/mL.	Catalog Number C009-60UL
DetectX[®] RBP Antibody A polyclonal antibody specific for RBP.	3 mL	Catalog Number C011-3ML
DetectX[®] RBP-Peroxidase Conjugate A RBP-peroxidase conjugate. in a special stabil		Catalog Number C010-5ML
<mark>Assay Buffer Concentrate</mark> A 5X concentrate that should be diluted with d		Catalog Number X053-28ML led water.
Wash Buffer Concentrate A 20X concentrate that should be diluted with		Catalog Number X007-30ML tilled water.
TMB Substrate	11 mL	Catalog Number X019-11ML
Stop Solution A 1M hydrochloric acid solution. Caustic.	5 mL	Catalog Number X020-5ML New Format
Plate Sealer	1 Each	Catalog Number X002-1EA

STORAGE INSTRUCTIONS

All components of this kit should be stored at 4 °C until the expiration date of the kit.



WEB INSERT OTHER MATERIALS REQUIRED

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Distilled or deionized water.

Repeater pipet with disposable tips capable of dispensing 25 μ L, 50 μ L and 100 μ L.

A microplate shaker and washer.

Colorimetric 96 well microplate reader capable of reading optical density at 450 nm.

Software for converting raw relative optical density readings from the plate reader and carrying out four parameter logistic curve (4PLC) fitting.

PRECAUTIONS

As with all such products, this kit should only be used by qualified personnel who have had laboratory safety instruction. The complete insert should be read and understood before attempting to use the product.

The antibody coated plate needs to be stored desiccated. The silica gel pack included in the foil ziploc bag will keep the plate dry. The silica gel pack will turn from blue to pink if the ziploc has not been closed properly.

The RBP Standard is purified from a human source and as such, should be treated as potentially hazardous. Proper safety procedures must be followed.

This kit utilizes a peroxidase-based readout system. Buffers, including other manufacturers Wash Buffers, containing sodium azide will inhibit color production from the enzyme. Make sure <u>all</u> buffers used for samples are **azide free**. Ensure that any plate washing system is rinsed well with deionized water prior to using the supplied Wash Buffer as prepared on Page 8.

The Stop Solution is acid. The solution should not come in contact with skin or eyes. Take appropriate precautions when handling this reagent.





WEB INSERT SAMPLE TYPES



This assay has been validated for human serum and EDTA and heparin plasma samples only. Samples containing visible particulate should be centrifuged prior to using. Moderate to severely hemolyzed samples should not be used in this kit. They have been shown to vield artificially high RBP concentrations.

For detecting RBP in urine samples please refer to the DetectX[®] Urinary Retinol Binding Protein Immunoassay kit, Catalog Number KU04-H1, that is 10 times more sensitive than this serum kit.

RBP is a highly conserved protein and we have shown that the Urinary RBP kit, which uses identical antibody and conjugate, will measure RBP's from sources other than human including rat, dog and rhesus monkey. The end user should evaluate recoveries of RBP in other plasma and serum samples being tested.

SAMPLE PREPARATION

Samples must be diluted 1:40 by taking one part of serum or plasma and adding thirtynine parts of diluted Assay Buffer (see page 8) prior to running in the kit. Any samples with RBP concentrations outside the standard curve range should be diluted further with Assav Buffer to obtain readings within the standard curve.

Use all samples within 2 hours of dilution.

REAGENT PREPARATION

We recommend that all standards and samples be run in duplicate to allow the end user to accurately determine RBP concentrations. Ensure that all samples have reached room temperature and have been diluted as appropriate prior to running them in the kit.

Assay Buffer New Format



Dilute Assay Buffer Concentrate 1:5 by adding one part of the concentrate to four parts of deionized water. Once diluted this is stable at 4°C for 3 months.

Wash Buffer

Dilute Wash Buffer Concentrate 1:20 by adding one part of the concentrate to nineteen parts of deionized water. Once diluted this is stable for 3 months at room temperature.



WEB INSERT

Standard Preparation - 10 µL Assay Format

For Normal RBP VAD Samples

Label test tubes as #1 through #8. Pipet 190 μ L of Assay Buffer into tube #1 and 100 μ L into tubes #2 to #8. Carefully add 10 μ L of the RBP stock solution to tube #1 and vortex completely. Take 100 μ L of the RBP solution in tube #1 and add it to tube #2 and vortex completely. Repeat the serial dilutions for tubes #3 through #8. The concentration of RBP in tubes 1 through 8 will be 5,000, 2,500, 1,250, 625, 312.5, 156.25, 78.125 and 39.06 ng/mL.

	Std 1	Std 2	Std 3	Std 4	Std 5	Std 6	Std 7	Std 8
Assay Buffer (µL)	190	100	100	100	100	100	100	100
Addition	Stock	Std 1	Std 2	Std 3	Std 4	Std 5	Std 6	Std 7
Vol of Addition (µL)	10	100	100	100	100	100	100	100
Final Conc (ng/mL)	5,000	2,500	1,250	625	312.5	156.25	78.125	39.06

Standard Preparation - 25 µL Assay Format

For Low RBP VAD Samples

Label test tubes as #1 through #8. Pipet 390 μ L of Assay Buffer into tube #1 and 100 μ L into tubes #2 to #8. Carefully add 10 μ L of the RBP stock solution to tube #1 and vortex completely. Take 100 μ L of the RBP solution in tube #1 and add it to tube #2 and vortex completely. Repeat the serial dilutions for tubes #3 through #8. The concentration of RBP in tubes 1 through 8 will be 2,500, 1,250, 625, 312.5, 156.25, 78.125, 39.06 and 19.53 ng/mL.

	Std 1	Std 2	Std 3	Std 4	Std 5	Std 6	Std 7	Std 8
Assay Buffer (µL)	390	100	100	100	100	100	100	100
Addition	Stock	Std 1	Std 2	Std 3	Std 4	Std 5	Std 6	Std 7
Vol of Addition (µL)	10	100	100	100	100	100	100	100
Final Conc (ng/mL)	2,500	1,250	625	312.5	156.25	78.125	39.06	19.53

Use all Standards within 2 hour of preparation.



WEB INSERT Assay Protocol - 10 µL and 25 µL

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- 1. Use the plate layout sheet on the back page to aid in proper sample and standard identification. Determine the number of wells to be used and return unused wells to the foil pouch with desiccant. Seal the ziploc plate bag and store at 4°C.
- 2. Pipet 10 μ L (25 μ L for alternate format) of samples or standards into wells in the plate.
- 3. Pipet 35 μ L (50 μ L for alternate format) of Assay Buffer into the non-specific binding (NSB) wells.
- 4. Pipet 10 μ L (25 μ L for alternate format) of Assay Buffer into wells to act as maximum binding wells (B0 or 0 ng/mL).
- 5. Add 50 µL of the DetectX[®] RBP Conjugate to each well using a repeater pipet.
- 6. Add 25 μ L of the DetectX[®] RBP Antibody to each well, **except the NSB wells**, using a repeater pipet.
- 7. Gently tap the sides of the plate to ensure adequate mixing of the reagents. Cover the plate with the plate sealer and shake at room temperature for 1 hour. If the plate is not shaken signals bound will be approximately 40% lower.
- Aspirate the plate and wash each well 4 times with 300 µL of Wash Buffer. Tap the plate dry on clean absorbent towels.
- 9. Add 100 μ L of the TMB Substrate to each well, using a repeater pipet.
- 10. Incubate the plate at room temperature for 30 minutes without shaking.
- 11. Add <mark>50 µL</mark> of the Stop Solution to each well, using a repeater or a multichannel pipet.
- 12. Read the optical density generated from each well in a plate reader capable of reading at 450 nm.
- 13. Use the plate reader's built-in 4PLC software capabilities to calculate RBP concentration for each sample.



WEB INSERT CALCULATION OF RESULTS



Average the duplicate OD readings for each standard and sample. Create a standard curve by reducing the data using computer software capable of generating a four-parameter logistic curve (4PLC) fit, after subtracting the mean OD's for the blank. The sample concentrations, calculated off the %B/BO curve, should be multiplied by the dilution factor to obtain neat sample values.

Or use the MyAssays online tool from <u>www.myassays.com/arbor-assays-retinol-binding-protein-</u> <u>serum-eia-kit.assay</u> to calculate the data.

MyAssays

QR code for Data Analysis:



*The MyAssays logo is a registered trademark of MyAssays Ltd.

		10	µL Assa	Ŋ	25 µL Assay			
Sample	Mean OD	Net OD	% B/B0	RBP Conc. (ng/mL)	Mean OD	Net OD	% B/B0	RBP Conc. (ng/mL)
NSB	0.110	0.000	-	-	0.106	0.000	-	-
Standard 1	0.361	0.251	11.5	5,000	0.306	0.200	10.6	2,500
Standard 2	0.492	0.382	17.5	2,500	0.430	0.324	17.2	1,250
Standard 3	0.662	0.552	25.2	1,250	0.574	0.468	24.9	625
Standard 4	0.903	0.793	36.2	625	0.763	0.657	34.9	312.5
Standard 5	1.169	1.059	48.4	312.5	0.978	0.872	46.4	156.3
Standard 6	1.463	1.353	61.8	156.3	1.217	1.111	59.1	78.125
Standard 7	1.744	1.634	74.6	78.125	1.500	1.394	74.1	39.06
Standard 8	1.935	1.825	83.4	39.06	1.675	1.569	83.5	19.53
во	2.299	2.189	100	0	1.986	1.880	100	0
Sample 1	0.857	0.747	34.1	698.5	0.535	0.429	22.8	721.6
Sample 2	1.588	1.478	67.5	115.3	1.097	0.991	52.7	114.3

TYPICAL DATA

Always run your own standard curve for calculation of results. Do not use this data.

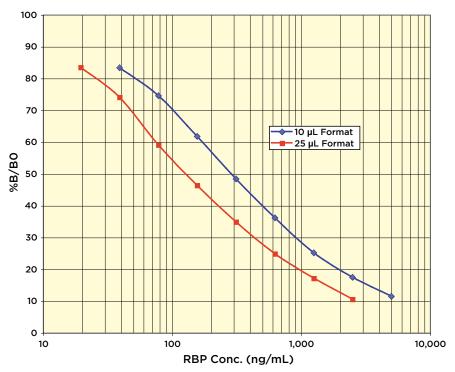
Conversion Factor: 1,000 ng/mL of human RBP is equivalent to 47.62 nM RBP.

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Typical Standard Curves



Always run your own standard curves for calculation of results. Do not use this data.

VALIDATION DATA

Sensitivity and Limit of Detection

Sensitivity was calculated by comparing the OD's for twenty wells run for each of the BO and standard #8. The sensitivity was determined at two (2) standard deviations from the BO along the standard curve.

10 µL Format Sensitivity was determined as 7.06 ng/mL.

25 µL Format Sensitivity was determined as 2.93 ng/mL.

The Limit of Detection was determined in a similar manner by comparing the OD's for twenty wells for each of the zero standard and a low concentration human sample.

10 μ L Format Limit of Detection was determined as 15.7 ng/mL.

25 μL Format Limit of Detection was determined as 5.26 ng/mL.

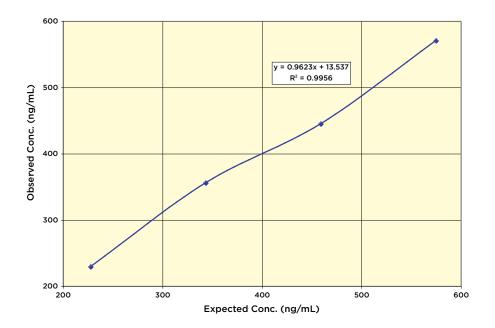


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Linearity

Linearity was determined in the 10 μL format using human serum and EDTA plasma samples, by taking samples with a high known RBP concentration and a lower RBP concentration and mixing them in the ratios given below. The measured RBP concentrations were compared to the expected values based on the ratios used.

High Sample	Low Sample	Expected Conc. Observed (ng/mL) Conc. (ng/mL)		% Recovery
80%	20%	575.0	570.3	99.2
60%	40%	459.3	444.8	96.8
40%	60%	343.6	355.6	103.5
20%	80%	228.0	228.9	100.4
			Mean Recovery	100.0%





Intra Assay Precision - 10 µL sample

Three human samples were diluted with Assay Buffer and run in replicates of 20 in an assay. The mean and precision of the calculated RBP concentrations were:

Sample	RBP Conc. (ng/mL)	%CV
1	614.2	5.6
2	227.1	8.2
3	90.8	12.9

Inter Assay Precision - 10 µL sample

Three human samples were diluted with Assay Buffer and run in duplicates in eighteen assays run over multiple days by three operators. The mean and precision of the calculated RBP concentrations were:

Sample	RBP Conc. (ng/mL)	%CV
1	681.0	6.7
2	261.1	8.0
3	112.1	9.7



WEB INSERT SAMPLE VALUES



Sixteen random human serum and plasma samples were tested in the assay. Values ranged from 23.2 to 35.2 μ g/mL with a average of 29.6 μ g/mL. The normal reference range for serum RBP is 30-60 μ g/mL¹⁰. Samples with a serum RBP level below about 15 μ g/mL RBP maybe considered Vitamin A deficient¹¹.

CROSS **R**EACTIVITY AND **I**NTERFERENTS

Two serum samples were spiked with varying concentrations of bilirubin, diluted 1:50 in Assay Buffer and tested in the assay. Bilirubin levels in normal serum are between 0.2 and 1.0 mg/dL¹². No significant change to the measured RBP levels were observed up to an additional 10.0 mg/dL of bilirubin.

A serum sample was spiked with varying concentrations of hemoglobin, diluted 1:50 in Assay Buffer and tested in the assay. No significant change to the measured RBP level was observed up to an additional 2 mg/mL of hemoglobin. However, moderately to severely hemolyzed samples should be avoided as they have been shown to yield artificially high RBP concentrations.

A serum sample was spiked with varying concentrations of lipids, diluted 1:40 in Assay Buffer and tested in the assay. No significant change to the measured RBP level was observed with the addition of high, medium and low levels of lipids.

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- 11. Hix J, Martinez C, Buchanan I, Morgan J, Tam M, Shankar A. "Development of a rapid enzyme immunoassay for the detection of retinol-binding protein." Am J Clin Nutr 2004;79:93-8.
- 12. Tietz, N.W., in "Textbook of Clinical Chemistry", WB Saunders, 1986.



WEB INSERT LIMITED WARRANTY



Arbor Assays warrants that at the time of shipment this product is free from defects in materials and workmanship. This warranty is in lieu of any other warranty expressed or implied, including but not limited to, any implied warranty of merchantability or fitness for a particular purpose.

We must be notified of any breach of this warranty within 48 hours of receipt of the product. No claim shall be honored if we are not notified within this time period, or if the product has been stored in any way other than outlined in this publication. The sole and exclusive remedy of the customer for any liability based upon this warranty is limited to the replacement of the product, or refund of the invoice price of the goods.

CONTACT INFORMATION

For details concerning this kit or to order any of our products please contact us:

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