ELSEVIER

The Spine Journal ■ (2008) ■

THE SPINE JOURNAL

Review Article

Contemporary concepts in spine care—the use of bone morphogenetic protein in spine fusion

Wellington K. Hsu, MD, Jeffrey C. Wang, MD*

Department of Orthopedic Surgery, and Rehabilitation, University of Wisconsin Madison, and the David Geffen School of Medicine, UCLA Comprehensive Spine Center, 1250 16th Street, 7th Floor Tower, Suite 745, Mail Code 703646, Santa Monica, CA 90404, USA

Received 14 January 2008; accepted 28 January 2008

Abstract

BACKGROUND CONTEXT: Because pseudarthrosis remains a clinically significant complication after spinal arthrodesis, the role of recombinant bone morphogenetic proteins (BMPs) is continually evaluated in spine surgery.

PURPOSE: This article reviews the important literature in clinical research involving the use of BMPs in the augmentation of spinal fusion.

STUDY DESIGN/SETTING: Review article.

METHODS: A literature search was performed via MEDLINE through PubMed with the dates January 1960 to July 2007 using the keywords "bone morphogenetic protein, BMP, spinal arthrodesis, and/or bone healing." Pertinent preclinical and clinical publications were chosen based on relevance and quality for inclusion in this study.

RESULTS: Publications focused on the historical context and potential clinical applications using BMP were selected to delineate the risks, benefits, and current indications for the augmentation of spinal arthrodesis.

CONCLUSIONS: Although multiple commercially available recombinant BMPs have demonstrated clinical success in interbody and posterolateral fusions, the associated costs preclude its routine use in spinal arthrodesis. The spine surgeon must assess each patient individually based on age, bone quality, diagnosis, comorbidities, and risks of nonunion to determine the cost effectiveness of the use of BMP to augment spinal fusion.

Keywords: BMP; Bone morphogenetic protein; Spine fusion; Spine arthrodesis

Introduction

Despite advances in the technologies and instrumentation of spine surgery, pseudarthrosis still occurs in 10% to 15% of all patients [1–4]. Furthermore, approximately 500,000 autogenous bone grafting procedures are performed annually, of which nearly 50% are used for spinal fusion [5]. Because the procurement of autologous bone graft is fraught with significant morbidity and postoperative pain [6–13], the study of bone graft substitutes in spine surgery has expanded to include recombinant growth factors, cell-based therapies, and the use of gene transfer strategies to enhance bone formation and improve fusion rates.

The significant rates of pseudarthroses and reports of operative morbidity from the harvest of autograft can limit the success rates of primary spine fusion in certain patients. In addition, the stringent biological environment created from revision procedures presents a more complicated array of problems and unpredictable outcomes after further surgical intervention. Dense fibrous tissue, intervertebral disc, and muscle cells commonly encountered during revision procedures have been found to inhibit host bone repair [14]. Because the success rates of fusion in this poor osteoinductive environment are relatively low, recent studies have been directed toward the development of new biologic substitutes to improve outcomes in both primary and revision procedures. For this reason, interest in bone graft substitutes and enhancers for the supplementation of spine surgery is on the rise.

FDA device/drug status: not applicable.

The author, JCW, acknowledges a financial relationship (Royalties from Medtronics, DePuy, Seaspine, Biomet; stock owner of Aesculp, Stryker; board of directors of K2M, Lanx/UCLA-Bone Biologics, Vertiflex) that may indirectly relate to the subject of this research.

The author, WKH, has no financial relationships to disclose.

* Corresponding author. UCLA Comprehensive Spine Center, Orthopedic Spine Service, David Geffen School of Medicine, 1250 16th Street, 7th Floor Tower, Suite 745, Mail Code 703646, Santa Monica, CA 90404, USA. Tel.: (310) 319-3334; fax: (310) 319-5055.

E-mail address: jwang@mednet.ucla.edu (J.C. Wang)

1529-9430/08/$ – see front matter
doi:10.1016/j.spinee.2008.01.008

Recombinant growth factors such as the bone morphogenetic proteins (BMPs) have been an important recent development in the armamentarium to enhance spinal arthrodesis rates. The significant osteoinductive potential of recombinant growth factors coupled with the avoidance of complications associated with bone graft harvest have encouraged the research into optimizing the clinical use of these powerful proteins. Although BMPs have a number of potential applications in spine surgery, this article will concentrate on the recent advances in the induction of spinal fusion.

Historical context and background

The discovery of BMPs by Urist in 1965 [15] has led to a diverse area of research dedicated to the identification and characterization of osteoinductive growth factors. Members of the Transforming Growth Factor (TGF)-ß superfamily, BMPs have been proposed for a number of applications in orthopedic surgery [16]. Although a total of 14 different BMPs have been reported [17], much of the recent study in the literature has focused on BMP-2, -6, -7, -9, and -14 (MP-52).

Recombinant BMP-2 (rhBMP-2) and BMP-7 (or osteogenic protein-1, rhOP-1) have been evaluated in numerous preclinical models, and successful healing in long bone defects has been reported [16,18–20]. Similar findings have been demonstrated in spinal arthrodesis models in animals [21–24]. US Food and Drug Administration (FDA) approval was recently granted for the use of rhBMP-2 to enhance anterior spinal fusion [25] and rhOP-1 to supplement posterior spine fusions [26]. In other orthopedic applications, human clinical trials evaluating the efficacy of rhBMP to treat open tibia fractures, distraction osteogenesis, and osteonecrosis of the hip are underway [27].

Clinical research

The efficacy of rhBMPs has been evaluated in preclinical models of spine fusion. Recombinant BMP-2 has been shown to reproducibly heal the lumbar spine in rodents and nonhuman primates [18,24,28–36]. Furthermore, rhOP-1 has also demonstrated consistent bone healing properties in rodent and sheep models [36–40]. Results from these studies suggest that the use of rhBMP results in similar if not superior fusion rates with biomechanically stronger fusion masses when compared with autogenous bone graft [18,24,28–36].

The first clinical pilot study using BMP in an anterior interbody fusion cage reported high rates of radiographic fusion with more rapid improvement in clinical outcome [25]. In a larger multicenter trial in 46 patients who underwent anterior lumbar discectemy and interbody fusion with cortical allograft dowels, the combination of rhBMP-2 on an absorbable collagen sponge was directly compared with autogenous iliac crest bone graft (ICBG) [41]. At the 12- and 24-month follow-up, patients who received rhBMP-2 had superior rates of fusion and improved clinical outcome determined by self-reporting questionnaires when compared with autogenous ICBG [41]. Moreover, the same investigators reported greater new bone formation outside the interbody fusion device with the use of rhBMP-2 when compared with the use of autograft [41]. These studies have subsequently led to FDA approval for the use of rhBMP-2 for human subjects in anterior spinal fusion. Since then, additional studies have expanded the potential clinical uses of rhBMPs in the spine.

Vaccaro et al. [42] recently demonstrated the efficacy of rhOP-1 putty (3.5 mg rhOP-1 with 1 g Type I collagen) in the enhancement of posterolateral lumbar arthrodesis. In a randomized, prospective, multicenter study, a total of 36 patients with degenerative spondylolisthesis were treated with either rhOP-1 or autogenous ICBG in an uninstrumented posterolateral fusion after a decompressive laminectemy. At 1-year follow-up, 74% (14 of 19 patients) of the rhOP-1 and 60% (6 of 10 patients) in the autograft groups achieved a successful clinical and radiographic posterolateral arthrodesis (Fig. 1), which was not statistically significant [42]. These authors concluded that fusion rates

Fig. 1. Lateral neutral (Left), flexion (Center), and extension (Right) radiographs of a patient treated with recombinant osteogenic protein-1 and autograft in a posterolateral spinal arthrodesis without instrumentation demonstrating radiographic fusion 12 months after surgical implantation (reprinted with permission from Vaccaro et al. Eur Spine J 2003;12:495–500).

in the absence of internal fixation with the use of rhOP-1 putty was safe and yielded comparable results to that of ICBG.

Similarly, Boden et al. [43] reported the successful clinical use of rhBMP-2 in the healing of a posterolateral spine fusion in a comparison study involving 25 patients. Clinical improvement as defined by the mean Oswestry Disability Index score (6 weeks postoperatively) was greatest in the rhBMP-2-treatment-only group. Interestingly, the authors concluded that the use of a higher dose of recombinant growth factor in nonhuman primates (1.5–2.0 mg/mL) than in rodents (0.2–0.4 mg/mL), was required in healing a posterolateral spine fusion [43]. To date, it remains unclear why concentrations of BMP a million times greater than that found in the human body are required to successfully induce a spinal arthrodesis [44–46].

Follow-up studies using rhBMP-2 have confirmed its successful use in inducing a posterolateral spinal fusion diagnosed by computed tomography scan [47,48]. Glassman et al. [48] reported the use of a large INFUSE kit (Medtronic, Memphis, TN) (12 mg rhBMP-2, 1.5 mg/mL) in the posterolateral fusion bed as equivalent fusion success to ICBG. The authors concluded that INFUSE can effectively substitute for ICBG for both one- and two-level posterolateral instrumented fusions. Dimar et al. published 2-year radiographic results from an FDA investigational device exemption study comparing ICBG and rhBMP-2 combined with a compression resistant matrix (CRM) carrier for single posterolateral fusions. The authors demonstrated that patients in the BMP/CRM group experienced significantly higher fusion rates yet had less surgical time and blood loss than the ICBG group [47].

Reports in preclinical models have also indicated that modifications in the carrier can enhance the efficacy and local delivery of BMP [49,50]. Previous research has shown the requirement of relatively high doses of BMP-2 when used with a Type I collagen sponge in animal models [22,51]. Commercially available INFUSE bone graft also currently uses an absorbable collagen sponge scaffold. One study has reported the advantages of carrier modifications involving the addition of fast-resorbing biphasic calcium phosphate granules or allograft chips [49]. These alterations have been shown to add significant compression resistance and increase local confinement of recombinant growth factor. Using these carrier modifications, the amount of rhBMP-2 required to heal a nonhuman primate spine model was decreased significantly (3 mg/side vs. up to 32 mg/side with plain collagen sponge). Recently, combined carriers consisting of bovine collagen and tricalcium/hydroxyapatite (CRM) have been used to deliver BMPs to the region of interest [47].

The importance of associated carriers with BMP was elucidated when Barnes et al. [52] reported the results of rhBMP-2 delivered on an absorbable collagen sponge wrapped around a bulking agent consisting of biphasic calcium phosphate and collagen in a posterolateral fusion model in rhesus monkeys. Results from this and other studies suggest that the required dosage of rhBMP-2 for spinal arthrodesis can be reduced by optimizing the delivery of growth factor by combining the strengths of different carriers [49]. Conversely, carriers such as fibrin glue have been shown to inhibit bone formation induced from rhBMP and may provide protection from heterotopic ossification and diffusion of protein to undesirable adjacent areas [53].

Because the treatment of spinal pseudarthrosis is fraught with relatively poor outcomes and potential complications, the interest in the utilization of rhBMP for these clinical challenges is on the rise. With the use of different preclinical pseudarthrosis models, recombinant growth factors may eventually prove to be a more appropriate bone graft option than other existing choices including ICBG. With the use of a nicotine-exposed rabbit lumbar pseudarthrosis model, Osteogenic Protein (OP)-1 was found to increase the expression of crucial genes in bone repair such as angiogenin, vascular endothelial growth factor, and BMPs [54]. In fact, these authors concluded that application of a single BMP in relatively high concentrations to a biologically stringent environment can induce angiogenic and osteogenic gene expression greater than that seen with autologous graft. A separate pseudarthrosis rabbit model exposed to preoperative radiation was used to demonstrate the superiority of rhBMP-2 to ICBG in producing a greater rate of fusion [55]. These studies are valuable in establishing the clinical and practical advantages in the use of recombinant growth factors for challenging biological environments. Not only can a more reliable osteoinductive stimulus be delivered to a fusion bed devoid of vascular supply and osteoinductivity, but the significant morbidities of autograft harvest can also be avoided.

Despite the overwhelming evidence in support of the routine use of rhBMPs in the enhancement of spinal arthrodesis, a number of studies have suggested potential complications with its clinical use. Smucker et al. [56] reported that 27.5% of a total of 69 patients who underwent anterior cervical spine fusions using rhBMP-2 had a clinical significant neck-swelling event compared with only 3.6% of patients in the non-rhBMP-2 group (Fig. 2). Other studies have confirmed the finding that the use of rhBMP-2 in the anterior cervical spine can be problematic [57,58]. Furthermore, the use of rhBMP-2 in transforaminal lumbar interbody fusion has been reported to lead to significant vertebral bone resorption in a total of 22 of 32 lumbar levels studied postoperatively with a computed tomography scan [59]. These authors concluded that rhBMP-2 was the direct cause of resorption, which led to graft subsidence and prevented solid radiographic union in a significant number of cases [59].

The continued research into the efficacy of rhBMPs in the augmentation of spinal arthrodesis offers promising results. Evidence from early clinical trials indicate that the use of rhBMPs results in fewer side effects, more rapid clinical improvement, and fusion rates that are as good

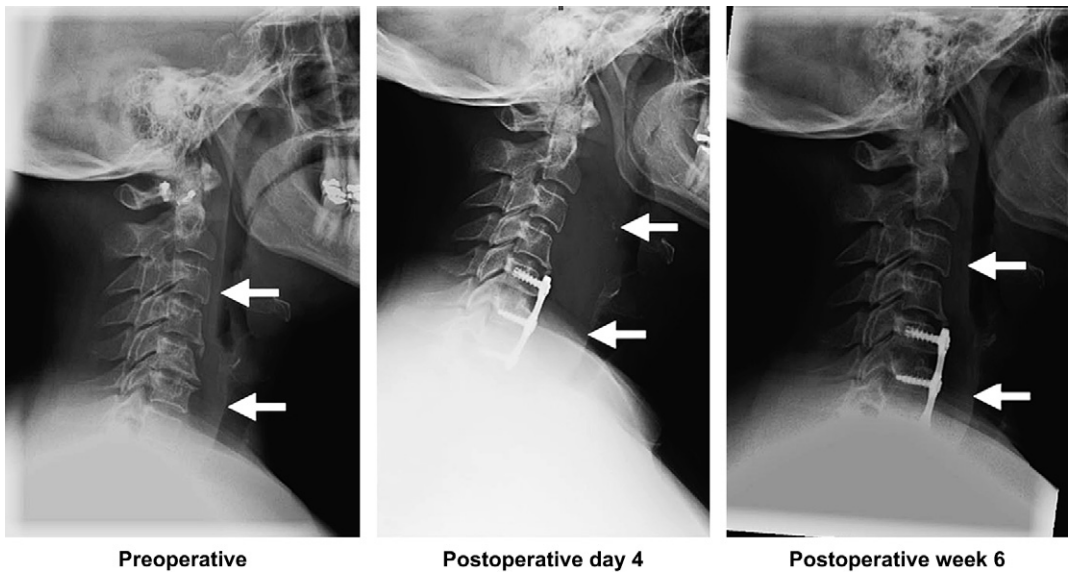

Fig. 2. Postoperative lateral cervical spine radiographs of a patient who underwent C5–7 anterior cervical discectomy and fusion with recombinant BMP-2, allograft, and plate. Arrows demonstrate marked severe anterior neck swelling and dysphagia on postoperative Day 4, which decreased to baseline by postoperative Week 6 (reprinted with permission from Smucker et al. Spine 2006;31(24): 2813–2819).

as, if not better than iliac crest autograft [25,39,41,43,47,60]. These studies have also shown that when used at an adequate dose with an appropriate carrier matrix, rhBMPs can be used for a wide range of spine problems with or without instrumentation for degenerative conditions, trauma, and even significant spinal deformity [61]. However, there remains great uncertainty with regard to the cost effectiveness of these recombinant growth factors, the potential burden on the health-care system from decades of recombinant protein use, and for which clinical indications they are most applicable. The identification of appropriate carriers for different clinical scenarios is even more essential to reduce both the dose and cost of recombinant proteins.

Discussion

Spinal arthrodesis is complicated by challenges including osteoporotic bone, a stringent biological environment and poor local vascularity. Bone graft substitutes are essential in obtaining consistent arthrodesis without the morbidity associated with autogenous graft. However, as a whole, available data vary widely regarding these substances and careful evaluation is necessary to identify the appropriate use of various bone graft agents. In making these critical decisions, surgeons must assess the host biologic environment and must ensure that four critical elements are present to promote bone repair: the presence of bioactive factors, responding cells, matrix, and an adequate vascular supply.

The body of evidence reporting the efficacy of rhBMP in clinical studies has grown considerably over the past 5 years. Since the first report of BMP-induced osteoinduction in a clinical trial [25], additional studies have reported the superiority of rhBMP-2 to the use of autogenous bone graft [25,39,41,43,47,60]. Moreover, patients treated with rhBMP-2 alone have been found to show more rapid and significant clinical improvement after spine fusion [43]. Reasons for this finding may be attributed to the subsequent shorter operative time and hospital stay [62]. Recent studies have shown that with adequate dosing of recombinant protein in patients without spinal instability, the use of rhBMP may decrease the need for instrumentation [42,63,64]. Furthermore, multiple investigators have demonstrated the osteoinductive versatility of rhBMP using multiple approaches. Recent evidence has reported the efficacy of rhBMP in posterolateral, interbody and transpedicular approaches in inducing radiographic and histologic spine fusion [39,41,43,59,60,64]. Future uses of rhBMP may lead to higher success rates in minimally invasive procedures and lessen surgical exposures and operative time.

However, despite excellent clinical results, many concerns still exist for the routine use of recombinant growth factors. Clinical studies that confirm the safety from the use of rhBMP-2 in humans [27,42] fall short in evaluating possible long-term effects. A number of complications have also been associated with its use in both the cervical and lumbar spine [56,57,59]. Furthermore, the cost of rhBMP currently precludes its routine use in spine arthrodesis, and further study will be necessary to delineate the clear indications in which BMPs should be used. At the current time, the administration of rhBMP to the surgical site requires a biological milieu that harbors responding cells and an adequate vascular supply to induce bone healing [65]. Stringent environments complicated by significant scar tissue in revision procedures, osteoporotic

bone stock, and multilevel constructs may require a more potent osteoinductive stimulus to provide a successful spine fusion.

Studies in our laboratory using a rat femoral defect and spinal arthrodesis model have indicated that a threshold level of BMP-2 production may be necessary to completely heal a large critical-sized defect [66–68]. The existence of a threshold level of BMP for osteogenic activity is an important finding because potential bone grafting strategies must provide an adequate osteoinductive signal not only of sufficient intensity but also of length to induce bone repair.

For these reasons, because of the exorbitant costs of the utilization of these recombinant proteins, its routine use is not recommended. However, individual patient characteristics that increase the risk of pseudarthrosis such as smoking, osteoporosis, multilevel and revision surgeries, and previous graft site harvest may justify the additional costs of BMPs as a bone graft substitute during surgery. Further studies delineating the indications of BMPs in spine surgery are warranted.

Future topics

The future of the use of bone graft substitutes to enhance spine arthrodesis remains bright. Apart from the proven efficacy of recombinant growth factors in inducing bone formation in the spine, new techniques are being developed to make the use of gene therapy systems more practical in delivering long-term BMPs in the spine [24,46,69,70]. Novel studies involving cell-based therapies may prove to offer a cost-effective option in bone repair using lower doses of rhBMP. Multiple avenues of research exist in the development of biologic substitutes for the enhancement of spine fusion. The continued laboratory and clinical characterization of spinal biologics will ultimately offer spine surgeons multiple options in the arena of spine fusion.

Conclusion with key points

- BMPs have demonstrated comparable fusion rates and clinical outcomes when compared with ICBG in both interbody and posterolateral fusions in prospective, randomized clinical studies.
- Continued research has focused on the optimization of a carrier for BMP to increase the delivery of protein to the region of interest.
- The utilization of BMPs has also been associated with unique complications such as local soft tissue edema and bone resorption.
- Because of the prohibitive cost of BMPs, its routine use in spine surgery is not necessarily recommended.
- Individual patient characteristics must be taken into account to justify its use to augment arthrodesis.
- New cell-based and gene therapies to increase the regional delivery of BMPs are under development to potentially offer a less expensive way to improve spine fusion rates.

References

[1] Bridwell KH, Sedgewick TA, O'Brien MF, Lenke LG, Baldus C. The role of fusion and instrumentation in the treatment of degenerative spondylolisthesis with spinal stenosis. J Spinal Disord 1993;6: 461–72.

[2] McGuire RA, Amundson GM. The use of primary internal fixation in spondylolisthesis. Spine 1993;18:1662–72.

[3] West JL 3rd, Bradford DS, Ogilvie JW. Results of spinal arthrodesis with pedicle screw-plate fixation. J Bone Joint Surg Am 1991;73: 1179–84.

[4] Zdeblick TA. A prospective, randomized study of lumbar fusion. Preliminary results. Spine 1993;18:983–91.

[5] Ludwig SC, Kowalski JM, Boden SD. Osteoinductive bone graft substitutes. Eur Spine J 2000;(9 Suppl 1):S119–25.

[6] Banwart JC, Asher MA, Hassanein RS. Iliac crest bone graft harvest donor site morbidity. A statistical evaluation. Spine 1995;20: 1055–60.

[7] Goulet JA, Senunas LE, DeSilva GL, Greenfield ML. Autogenous iliac crest bone graft. Complications and functional assessment. Clin Orthop 1997;339:76–81.

[8] Reid RL. Hernia through an iliac bone-graft donor site. A case report. J Bone Joint Surg Am 1968;50:757–60.

[9] Coventry MB, Tapper EM. Pelvic instability: a consequence of removing iliac bone for grafting. J Bone Joint Surg Am 1972;54: 83–101.

[10] Challis JH, Lyttle JA, Stuart AE. Strangulated lumbar hernia and volvulus following removal of iliac crest bone graft. Acta Orthop Scand 1975;46:230–3.

[11] Cowley SP, Anderson LD. Hernias through donor sites for iliac-bone grafts. J Bone Joint Surg Am 1983;65:1023–5.

[12] Kuhn DA, Moreland MS. Complications following iliac crest bone grafting. Clin Orthop 1986;224–6.

[13] Kurz LT, Garfin SR, Booth RE Jr. Harvesting autogenous iliac bone grafts. A review of complications and techniques. Spine 1989;14: 1324–31.

[14] Bae H, Kanim LEA, Zhao L, Wong P, Delamarter R. Cellular environments alter performance of rhBMP-2 and induce pseudarthrosis. Spine J 2004;4(5S):52S.

[15] Urist MR. Bone: formation by autoinduction. Science 1965;150: 893–9.

[16] Zabka AG, Pluhar GE, Edwards RB 3rd, et al. Histomorphometric description of allograft bone remodeling and union in a canine segmental femoral defect model: a comparison of rhBMP-2, cancellous bone graft, and absorbable collagen sponge. J Orthop Res 2001;19: 318–27.

[17] Cheng H, Jiang W, Phillips FM, et al. Osteogenic activity of the fourteen types of human bone morphogenetic proteins (BMPs). J Bone Joint Surg Am 2003;85-A:1544–52.

[18] Cook SD, Dalton JE, Tan EH, Whitecloud TS 3rd, Rueger DC. In vivo evaluation of recombinant human osteogenic protein (rhOP-1) implants as a bone graft substitute for spinal fusions. Spine 1994; 19:1655–63.

[19] Cook SD, Wolfe MW, Salkeld SL, Rueger DC. Effect of recombinant human osteogenic protein-1 on healing of segmental defects in nonhuman primates. J Bone Joint Surg Am 1995;77:734–50.

[20] Yasko AW, Lane JM, Fellinger EJ, Rosen V, Wozney JM, Wang EA. The healing of segmental bone defects, induced by recombinant human bone morphogenetic protein (rhBMP-2). A radiographic, histological, and biomechanical study in rats. J Bone Joint Surg Am 1992;74:659–70.

[21] Boden SD, Martin GJ Jr, Horton WC, Truss TL, Sandhu HS. Laparoscopic anterior spinal arthrodesis with rhBMP-2 in a titanium interbody threaded cage. J Spinal Disord 1998;11:95–101.
[22] Martin GJ Jr, Boden SD, Marone MA, Moskovitz PA. Posterolateral intertransverse process spinal arthrodesis with rhBMP-2 in a nonhuman primate: important lessons learned regarding dose, carrier, and safety. J Spinal Disord 1999;12:179–86.
[23] Sandhu HS, Kanim LE, Toth JM, et al. Experimental spinal fusion with recombinant human bone morphogenetic protein-2 without decortication of osseous elements. Spine 1997;22: 1171–80.
[24] Wang JC, Kanim LE, Yoo S, Campbell PA, Berk AJ, Lieberman JR. Effect of regional gene therapy with bone morphogenetic protein-2-producing bone marrow cells on spinal fusion in rats. J Bone Joint Surg Am 2003;85-A:905–11.
[25] Boden SD, Zdeblick TA, Sandhu HS, Heim SE. The use of rhBMP-2 in interbody fusion cages. Definitive evidence of osteoinduction in humans: a preliminary report. Spine 2000;25:376–81.
[26] Available at http://www.fda.gov/cdrh/pdf2/H020008a.pdf. Accessed on 12/1/07.
[27] Valentin-Opran A, Wozney J, Csimma C, Lilly L, Riedel GE. Clinical evaluation of recombinant human bone morphogenetic protein-2. Clin Orthop 2002;395:110–20.
[28] Boden SD, Moskovitz PA, Morone MA, Toribitake Y. Video-assisted lateral intertransverse process arthrodesis. Validation of a new minimally invasive lumbar spinal fusion technique in the rabbit and nonhuman primate (rhesus) models. Spine 1996;21:2689–97.
[29] Holliger EH, Trawick RH, Boden SD, Hutton WC. Morphology of the lumbar intertransverse process fusion mass in the rabbit model: a comparison between two bone graft materials—rhBMP-2 and autograft. J Spinal Disord 1996;9:125–8.
[30] Martin GJ Jr, Boden SD, Titus L. Recombinant human bone morphogenetic protein-2 overcomes the inhibitory effect of ketorolac, a nonsteroidal anti-inflammatory drug (NSAID), on posterolateral lumbar intertransverse process spine fusion. Spine 1999;24:2188–93; discussion 2193–4.
[31] Schimandle JH, Boden SD, Hutton WC. Experimental spinal fusion with recombinant human bone morphogenetic protein-2. Spine 1995;20:1326–37.
[32] Silcox DH 3rd, Boden SD, Schimandle JH, Johnson P, Whitesides TE, Hutton WC. Reversing the inhibitory effect of nicotine on spinal fusion using an osteoinductive protein extract. Spine 1998;23:291–6; discussion 297.
[33] Boden SD, Martin GJ Jr, Morone M, Ugbo JL, Titus L, Hutton WC. The use of coralline hydroxyapatite with bone marrow, autogenous bone graft, or osteoinductive bone protein extract for posterolateral lumbar spine fusion. Spine 1999;24:320–7.
[34] Muschler GF, Hyodo A, Manning T, Kambic H, Easley K. Evaluation of human bone morphogenetic protein 2 in a canine spinal fusion model. Clin Orthop 1994;308:229–40.
[35] Sandhu HS, Kanim LE, Kabo JM, et al. Evaluation of rhBMP-2 with an OPLA carrier in a canine posterolateral (transverse process) spinal fusion model. Spine 1995;20:2669–82.
[36] Grauer JN, Patel TC, Erulkar JS, Troiano NW, Panjabi MM, Friedlaender GE. 2000 Young Investigator Research Award winner. Evaluation of OP-1 as a graft substitute for intertransverse process lumbar fusion. Spine 2001;26:127–33.
[37] Kalodiki EP, Hoppensteadt DA, Nicolaides AN, et al. Deep venous thrombosis prophylaxis with low molecular weight heparin and elastic compression in patients having total hip replacement. A randomised controlled trial. Int Angiol 1996;15:162–8.
[38] Magin MN, Delling G. Improved lumbar vertebral interbody fusion using rhOP-1: a comparison of autogenous bone graft, bovine hydroxylapatite (Bio-Oss), and BMP-7 (rhOP-1) in sheep. Spine 2001;26:469–78.
[39] Blattert TR, Delling G, Dalal PS, Toth CA, Balling H, Weckbach A. Successful transpedicular lumbar interbody fusion by means of a composite of osteogenic protein-1 (rhBMP-7) and hydroxyapatite carrier: a comparison with autograft and hydroxyapatite in the sheep spine. Spine 2002;27:2697–705.
[40] Masuda K, Takegami K, An H, et al. Recombinant osteogenic protein-1 upregulates extracellular matrix metabolism by rabbit annulus fibrosus and nucleus pulposus cells cultured in alginate beads. J Orthop Res 2003;21:922–30.
[41] Burkus JK, Gornet MF, Dickman CA, Zdeblick TA. Anterior lumbar interbody fusion using rhBMP-2 with tapered interbody cages. J Spinal Disord Tech 2002;15:337–49.
[42] Vaccaro AR, Patel T, Fischgrund J, et al. A pilot safety and efficacy study of OP-1 putty (rhBMP-7) as an adjunct to iliac crest autograft in posterolateral lumbar fusions. Eur Spine J 2003;12:495–500.
[43] Suh DY, Boden SD, Louis-Ugbo J, et al. Delivery of recombinant human bone morphogenetic protein-2 using a compression-resistant matrix in posterolateral spine fusion in the rabbit and in the nonhuman primate. Spine 2002;27:353–60.
[44] Baltzer AW, Lieberman JR. Regional gene therapy to enhance bone repair. Gene Ther 2004;11:344–50.
[45] Hsu W, Sugiyama O, Feeley B, et al. Lentiviral-mediated BMP-2 gene transfer enhances healing of segmental femoral defects in rats. Poster presentation at American Society of Bone and Mineral Research annual meeting; 2004.
[46] Hsu WK, Sugiyama O, Park SH, et al. Lentiviral-mediated BMP-2 gene transfer enhances healing of segmental femoral defects in rats. Bone 2007;40:931–8.
[47] Dimar JR, Glassman SD, Burkus KJ, Carreon LY. Clinical outcomes and fusion success at 2 years of single-level instrumented posterolateral fusions with recombinant human bone morphogenetic protein-2/compression resistant matrix versus iliac crest bone graft. Spine 2006;31:2534–9; discussion 2540.
[48] Glassman SD, Carreon L, Djurasovic M, et al. Posterolateral lumbar spine fusion with INFUSE bone graft. Spine J 2007;7:44–9.
[49] Akamaru T, Suh D, Boden SD, Kim HS, Minamide A, Louis-Ugbo J. Simple carrier matrix modifications can enhance delivery of recombinant human bone morphogenetic protein-2 for posterolateral spine fusion. Spine 2003;28:429–34.
[50] Liao SS, Guan K, Cui FZ, Shi SS, Sun TS. Lumbar spinal fusion with a mineralized collagen matrix and rhBMP-2 in a rabbit model. Spine 2003;28:1954–60.
[51] Boden SD, Schimandle JH, Hutton WC, Chen MI. 1995 Volvo Award in basic sciences. The use of an osteoinductive growth factor for lumbar spinal fusion. Part I: Biology of spinal fusion. Spine 1995;20: 2626–32.
[52] Barnes B, Boden SD, Louis-Ugbo J, et al. Lower dose of rhBMP-2 achieves spine fusion when combined with an osteoconductive bulking agent in non-human primates. Spine 2005;30:1127–33.
[53] Patel VV, Zhao L, Wong P, et al. Controlling bone morphogenetic protein diffusion and bone morphogenetic protein-stimulated bone growth using fibrin glue. Spine 2006;31:1201–6.
[54] White AP, Maak TG, Prince D, et al. Osteogenic protein-1 induced gene expression: evaluation in a posterolateral spinal pseudarthrosis model. Spine 2006;31:2550–5.
[55] Ames CP, Smith JS, Preul MC, et al. Effect of recombinant human bone morphogenetic protein-2 in an experimental model of spinal fusion in a radiated area. Spine 2005;30:2585–92.
[56] Smucker JD, Rhee JM, Singh K, Yoon ST, Heller JG. Increased swelling complications associated with off-label usage of rhBMP-2 in the anterior cervical spine. Spine 2006;31:2813–9.
[57] Shields LB, Raque GH, Glassman SD, et al. Adverse effects associated with high-dose recombinant human bone morphogenetic protein-2 use in anterior cervical spine fusion. Spine 2006;31:542–7.
[58] Perri B, Cooper M, Lauryssen C, Anand N. Adverse swelling associated with use of rh-BMP-2 in anterior cervical discectomy and fusion: a case study. Spine J 2007;7:235–9.
[59] McClellan JW, Mulconrey DS, Forbes RJ, Fullmer N. Vertebral bone resorption after transforaminal lumbar interbody fusion with bone

morphogenetic protein (rhBMP-2). J Spinal Disord Tech 2006;19: 483–6.

[60] Vaccaro AR, Anderson DG, Patel T, et al. Comparison of OP-1 Putty (rhBMP-7) to iliac crest autograft for posterolateral lumbar arthrodesis: a minimum 2-year follow-up pilot study. Spine 2005;30:2709–16.

[61] Luhmann SJ, Bridwell KH, Cheng I, Imamura T, Lenke LG, Schootman M. Use of bone morphogenetic protein-2 for adult spinal deformity. Spine 2005;30(17 Suppl):S110–7.

[62] Vaccaro AR, Chiba K, Heller JG, et al. Bone grafting alternatives in spinal surgery. Spine J 2002;2:206–15.

[63] Jenis LG, Wheeler D, Parazin SJ, Connolly RJ. The effect of osteogenic protein-1 in instrumented and noninstrumented posterolateral fusion in rabbits. Spine J 2002;2:173–8.

[64] Boden SD, Kang J, Sandhu H, Heller JG. Use of recombinant human bone morphogenetic protein-2 to achieve posterolateral lumbar spine fusion in humans: a prospective, randomized clinical pilot trial: 2002 Volvo Award in clinical studies. Spine 2002;27:2662–73.

[65] Wang JC, Kanim LE, Nagakawa IS, Yamane BH, Vinters HV, Dawson EG. Dose-dependent toxicity of a commercially available demineralized bone matrix material. Spine 2001;26:1429–35; discussion 1435–6.

[66] Hsu W, Wang J, Feeley B, et al. Gene therapy utilizing stem cells from human fat in a rat posterolateral spine fusion model. Spine J 2004;4(5S):51S.

[67] Lieberman JR, Daluiski A, Stevenson S, et al. The effect of regional gene therapy with bone morphogenetic protein-2-producing bone-marrow cells on the repair of segmental femoral defects in rats. J Bone Joint Surg Am 1999;81:905–17.

[68] Feeley BT, Krenek L, Liu N, et al. Overexpression of noggin inhibits BMP-mediated growth of osteolytic prostate cancer lesions. Bone 2006;38:154–66.

[69] Lieberman JR, Le LQ, Wu L, et al. Regional gene therapy with a BMP-2-producing murine stromal cell line induces heterotopic and orthotopic bone formation in rodents. J Orthop Res 1998;16: 330–9.

[70] Peterson B, Zhang J, Iglesias R, et al. Healing of critically sized femoral defects, using genetically modified mesenchymal stem cells from human adipose tissue. Tissue Eng 2005;11:120–9.