



CSP LOW-RISK LEVEL MEDIA-FILL CHALLENGE KIT

Cat. no. HVL1	Low-Risk Level Media-Fill Challenge Test Kit	Single-Use Kit
Each kit contains:		
	Tryptic Soy Broth (TSB), 100ml Serum Vial, 100ml	1 vial
	Sterile 20ml Serum Vials (empty)	3 vials
	Whirl-Pak® Bag	1 bag
	Results Log Sheet	1 sheet

INTENDED USE

CSP Low-Risk Level Media-Fill Challenge Test Kit is recommended for routine use in the monitoring of aseptic procedures used in Compounding Sterile Preparations (CSPs).

The kit contains the necessary materials for one pharmacist or technician to perform the low-risk level media-fill challenge testing as specified in USP Chapter <797>.

SUMMARY

On January 1, 2004 Chapter <797>, of the United States Pharmacopeia/National Formulary USP27/NF22) entitled "Pharmaceutical Compounding Sterile Preparations," became effective. USP Chapter <797> details the procedures and requirements for compounding sterile preparations and sets standards that are applicable to all practice settings in which sterile preparations are compounded. Since USP Chapter <797> is considered a requirement, pharmacies may be subject to inspection for compliance with these standards by State Boards of Pharmacy, the FDA, and accreditation organizations, such as the Joint Commission, Accreditation Commission for Health Care, Inc. (ACHA) and the Community Health Accreditation Program (CHAP). Compliance with these standards was required by January 1, 2006.

USP Chapter <797> defines three levels of risk related to sterile preparation and includes quality assurance requirements for each risk level. These risk levels are based on the potential for introducing sources of contamination to the preparations from microbial, chemical or physical contamination during compounding activities, or in the case of high-risk compounding that the product would remain contaminated. USP Chapter <797> provides general guidance on risk level assignment based upon compounding manipulations, types of ingredients and equipment used, compounding environment, and storage and use of the resulting preparation. A summary table is included, but regardless of the examples provided, *the ultimate determination of risk level is the responsibility of the licensed health care professionals who supervise compounding, using their professional judgement and experience.*

Risk Level	Description	Examples	QA Monitoring and Frequency
Low	<ul style="list-style-type: none"> Involves only transfer, measuring and mixing with closed or sealed packaging systems. Limited to aseptically opening ampules, penetrating sterile stoppers on vials with sterile needles or syringes, transferring sterile liquids in sterile syringes to sterile administration devices. Prepared entirely in an ISO Class 5 (see below) or better air quality environment. In the absence of passing a sterility test, storage for a maximum of 48 hours at room temperature, 14 days refrigerated, or 45 days in solid frozen state. 	<ul style="list-style-type: none"> Single transfers of sterile dosage forms from ampules, bottles, bags, and vials using sterile syringes with sterile needles. (<i>Vancomycin 1gm in NS 100ml prepared for 1 patient.</i>) Manually measuring and mixing no more than 3 manufactured products to compound drug admixtures and nutritional solutions. (<i>TPN solution compounded using gravity transfer of commercially available sterile Amino Acid and Dextrose solutions and no more than 3 sterile additives transferred using a syringe and needles.</i>) 	<p>Media-Fill Challenge Test <i>Low-risk Media-fill Challenge</i> <u>Hardy Cat. no.:</u> HVL1 <u>Frequency:</u> Annual testing for each person who compounds low-risk sterile preparations.</p> <p>Environmental monitoring: <i>Air Monitoring:</i> <u>Hardy Cat. no.:</u> G60 <u>Frequency:</u> At least semi-annual testing of each Laminar Air Flow Workbench or barrier isolator.</p> <p><i>Surface and Glove Fingertips:</i> <u>Hardy Cat. no.:</u> P34 <u>Frequency:</u> Surface monitoring required on a periodic basis of each Laminar Air Flow Workbench or barrier isolator. Glove fingertips shall be done at initial competency evaluation and no less than three times before being allowed to compound sterile preparations. Re-evaluation is required at each media fill challenge test.</p>
Medium	<ul style="list-style-type: none"> Multiple individual or small doses are combined or pooled to prepare a CSP for administration to multiple patients or to one patient on multiple occasions. Involves complex aseptic manipulations or requires a long duration to prepare. Does not contain broad-spectrum bacteriostatic substances, and is administered over several days (e.g. worn or implanted infusion device). Prepared entirely in an ISO Class 5 (see below) or better air quality environment. In the absence of passing a sterility test, storage for a maximum of 30 hours at room temperature, 9 days refrigerated, or 45 days at solid frozen state. 	<ul style="list-style-type: none"> TPN fluids using manually or automated compounded, involving multiple injections, detachments, and attachments of nutrient source products to deliver components to a final sterile container. Filling reservoirs of injection and infusion devices with multiple sterile drug products and evacuation of air from those reservoirs before dispensing. (<i>Chemotherapy prepared for infusion over 5 days using a portable infusion device.</i>) Filling reservoirs of injection and infusion devices with sterile drug solutions that will be administered over several days at ambient temperatures between 25° and 40°C. (<i>Implanted pump reservoir filled with Preservative-Free Morphine for infusion over 4 weeks.</i>) Transfer from multiple ampules or vials into a single, final sterile container or product. (<i>Any IV solution compounded with more than 3 additives.</i>) 	<p>Media-Fill Challenge Test <i>Medium-risk Media-fill Challenge</i> <u>Hardy Cat. no.:</u> HVM1 <u>Frequency:</u> Annual testing for each person who compounds medium-risk sterile preparations.</p> <p>Environmental Monitoring: <i>Air Monitoring:</i> <u>Hardy Cat. no.:</u> G60 <u>Frequency:</u> At least semi-annual testing of each Laminar Air Flow Workbench or barrier isolator.</p> <p><i>Surface and Glove Fingertips:</i> <u>Hardy Cat. no.:</u> P34 <u>Frequency:</u> Surface monitoring required on a periodic basis of each Laminar Air Flow Workbench or barrier isolator. Glove fingertips shall be done at initial competency evaluation and no less than three times before being allowed to compound sterile preparations. Re-evaluation is required at each media fill challenge test.</p>
High	<ul style="list-style-type: none"> Non-sterile ingredients are incorporated or a non-sterile device is employed before terminal sterilization. Non-sterile ingredients are incorporated, or a non-sterile device is employed before terminal sterilization. Non-sterile components are exposed for at least 6 hours before being sterilized. Exposed to air quality inferior to ISO Class 5 (see below). In the absence of passing a sterility test, storage for a maximum of 24 hours at room temperature, 3 days refrigerated, or 45 days in solid frozen state. 	<ul style="list-style-type: none"> Dissolving non-sterile bulk drug and nutrient powders to make solutions which will be terminally sterilized. (<i>TPN solutions made from dry amino acids.</i>) Measuring and mixing sterile ingredients in non-sterile devices before sterilization is performed. (<i>Ophthalmic solution filtered into a non-sterile dropper bottle.</i>) 	<p>Media-Fill Challenge Test <i>High-risk Media-fill Challenge</i> <u>Hardy Cat. no.:</u> HVH1 <u>Frequency:</u> Semi-annual for each person who compounds high-risk sterile preparations.</p> <p>Environmental: <i>Air Monitoring:</i> <u>Hardy Cat. no.:</u> G60 and W28 <u>Frequency:</u> At least semi-annual testing of each Laminar Air Flow Workbench or barrier isolator.</p> <p><i>Surface and Glove Fingertips:</i> <u>Hardy Cat. no.:</u> P34 <u>Frequency:</u> Surface monitoring required on a periodic basis of each Laminar Air Flow Workbench or barrier isolator. Glove fingertips shall be done at initial competency evaluation and no less than three times before being allowed to compound sterile preparations. Re-evaluation is required at each media fill challenge test.</p>

Source of risk level information: www.uspnf.com

Training for personnel who compound sterile preparations is mandatory and should be comprehensive and include thorough evaluation. Media-fill challenge testing (media-fill verification of technique) is used to verify that personnel have the necessary skills to compound aseptic preparations. During media-fill challenge testing, personnel are instructed to prepare a CSP using sterile liquid culture medium. The resulting solution is then incubated at 25-35 degrees C for 14 days. The solution is examined for the evidence of microbial growth or turbidity, during incubation and at the end of the 14 day period. If there is evidence of turbidity, the challenge test has failed and it can be concluded that there was a breach in aseptic technique.

All personnel who compound CSPs must complete media-fill challenge testing before they are allowed to compound CSPs. All personnel who compound CSPs must complete media-fill challenge testing before they are allowed to compound CSPs. As specified in the USP Chapter <797>, the testing should take place at least annually for low- and medium-risk CSPs and semi-annually for high-risk CSPs.

Tryptic Soy Broth, the medium used in the media-fill challenge testing, is widely used for the cultivation of microorganisms from environmental sources supporting the growth of the majority of bacteria and fungi. Tryptic Soy Broth, also known as TSB or Soybean-Casein Digest Broth, conforms to the formula given by the U.S. Pharmacopeia.⁽¹⁾ This medium contains digests of soybean meal and casein, which provide amino acids and other nitrogenous substances, making it a highly nutritious medium for a variety of organisms. Sodium chloride is added to maintain the osmotic equilibrium. Dextrose is incorporated as an energy source. The dipotassium phosphate is included in the formulation as a buffer to maintain the proper pH.

FORMULA

Ingredients per liter of deionized water:*

Pancreatic Digest of Casein	17.0gm
Sodium Chloride	5.0gm
Papaic Digest of Soybean Meal	3.0gm
Dextrose	2.5gm
Dipotassium Phosphate	2.5gm

Final pH 7.3 +/- 0.2 at 25 degrees C

*Adjusted and/or supplemented as required to meet performance criteria.

STORAGE AND SHELF LIFE

Storage: Upon receipt, store at 2-30 degrees C away from direct light. Media should not be used if there are any signs of deterioration (discoloration), contamination, or if the expiration date has passed. Protect from light, excessive heat, moisture, and freezing.

The expiration date applies to the product in its intact packaging when stored as directed.

This product has the following shelf life from the date of manufacture:

365 Days: HVL1 HardyVal™ Low-Risk Level Media-Fill Challenge Kit

Refer to the keyword "Storage," in the Hardy Diagnostics software program HUGO™, for more information on storing culture media.

PRECAUTIONS

For laboratory use only. Not for injection or ingestion. Observe approved biohazard precautions and aseptic techniques. This product is to be used only by adequately trained and qualified personnel. Sterilize all biohazard waste before disposal.

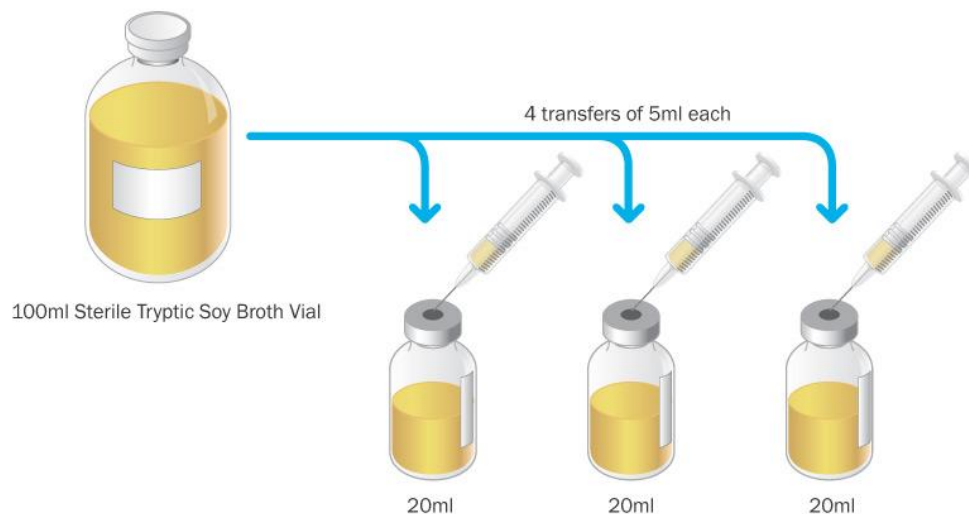
Refer to the keyword "Precautions," in the Hardy Diagnostics software program HUGO™, for more information regarding general precautions when using culture media.

Refer to the keyword "MSDS," in the Hardy Diagnostics software program HUGO™, for more information on handling potentially hazardous material.

PROCEDURE

When performing media-fill risk challenges, use procedures and techniques that most closely resemble those used during routine compounding of Low-Risk Level CSPs. If necessary, the following procedure may be modified to include more complex manipulations. Once begun, the test is completed without interruption.

1. Label the empty sterile 20ml vials with the date challenge is performed and the initials of the person performing the procedure.
2. Within an ISO Class 5 air quality environment, using the same sterile 10ml syringe and vented needle, transfer four 5ml aliquots of sterile Tryptic Soy Broth into the separate empty vials (total volume in each vial will be 20ml).
3. Apply sterile adhesive seals aseptically to the rubber closures once the vials have been filled.
4. Place the vials in a Whirl-Pak® bag for transport to the incubator.
5. Incubate the vials at 20 to 25 and/or 30 to 35 degrees C for 14 days. If two temperatures of incubation are selected, incubate for 7 days at each temperature.
6. Examine for the presence of turbidity. **Note:** Growth may not be evenly dispersed throughout the vial. Tap or swirl the vial to observe for growth that may have settled at the bottom. (If growth is observed the vial may be discarded. Do not continue to incubate for the full 14 days.)
7. Record results on the "Results Log Sheet".
8. Discard all used syringes, needles and completed test vials as biomedical waste.



INTERPRETATION OF RESULTS

Visible growth or turbidity seen on or before 14 days of incubation is a positive test for the presence of bacteria. The media-fill challenge test has failed and indicates that a non-sterile technique was used during the test.

No growth or turbidity observed in 14 days indicates the media-fill challenge test was successful and technique used during the media-fill challenge test was aseptic.

LIMITATIONS

Rare fastidious organisms may not grow in Tryptic Soy Broth.

Refer to the keyword "Limitations," in the Hardy Diagnostics software program HUGO™, for more information regarding general limitations on culture media.

MATERIALS REQUIRED BUT NOT PROVIDED

Standard supplies and equipment such as syringes, needles, adhesive seals, thermometers, incinerators, incubators, etc., as well as serological and biochemical reagents are not provided.

QUALITY CONTROL

The following organisms are routinely used for testing at Hardy Diagnostics:

Test Organisms	Inoculation Method*	Incubation			Results
		Time	Temperature	Atmosphere	
<i>Bacillus subtilis</i> ATCC® 6633	J	1-3 days	25-35°C	Aerobic	Growth; turbidity
<i>Candida albicans</i> ATCC® 10231	J	1-5 days	25-35°C	Aerobic	Growth; turbidity
<i>Aspergillus brasiliensis</i> formerly <i>A. niger</i> ATCC® 16404	J	1-5 days	25-35°C	Aerobic	Growth; turbidity

USER QUALITY CONTROL

Check for signs of contamination and deterioration. Users of commercially prepared media may be required to perform quality control testing with at least one known organism to demonstrate growth or a positive reaction; and at least one organism to demonstrate inhibition or a negative reaction (where applicable). Refer to the following keywords in HUGO™ for more information on QC: "Introduction to QC" and "QC of Finished Product".

* Refer to the keyword "Inoculation Procedures", in the Hardy Diagnostics software program HUGO™, for a description of inoculation procedures.

PHYSICAL APPEARANCE

Tryptic Soy Broth should appear clear and light amber in color.

REFERENCES

1. USP General Chapter <797> Pharmaceutical Compounding - Sterile Preparations. 2004. *USP27-22*. United States Pharmacopeial Convention, Rockville, MD.
2. USP General Chapter <797> Pharmaceutical Compounding - Sterile Preparations. *USP27-22: 1st Supplement*, April 2004. United States Pharmacopeial Convention, Rockville, MD.
3. www.uspnf.com
4. USP General Chapter <797> Pharmaceutical Compounding – Sterile Preparations Final Revisions. 2007. P. 1-61. United States Pharmacopeial Convention, Inc. Rockville, MD.
www.usp/pdf/EN/USPNF/generalChapter797.pdf
5. USP <797> Guidebook to Pharmaceutical Compounding – Sterile Preparations 2008 The United States Pharmacopeial Convention, Rockville, MD.

ATCC is a registered trademark of the American Type Culture Collection.

Whirl-Pak is a registered trademark of Nasco Industries, Inc.

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LOG SHEET

LOW-RISK LEVEL MEDIA-FILL CHALLENGE

Name: _____

Date Test Performed: _____

Signature: _____

Kit Lot #: _____

Kit Expiration Date: _____

Vial Number	Hood Number	Incubation Temp. ¹	Length of Incubation ²	Result: Growth/No growth	Interpretation: Pass/Fail	Notes/Corrective Action (Attach additional pages if necessary)
1						
2						
3						

¹ Recommended incubation temperature is 20 to 25 and/or 30 to 35°C per USP <797>.

² Recommended length of incubation is 14 days for negative cultures.

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Supervisor Signature

Date