

Import Export Data in R.

scan: Basic function to input data from a text file, **scan** returns a vector of numbers, strings , or logical.

To read from the console use: `scan()`

To read from a file use: `scan(filepath, what = " ")`

Example:

```
> z <- scan()
1: 1 2
3: 4 6 7
6: 10 11
8:
Read 7 items
> z
[1] 1 2 4 6 7 10 11
```

read.table: To read data from a text file in the form of a data table and produce a data frame in R.

`read.table(file, sep=" ",head=F)`

read.csv: To read data from a “,” delimited file in the form of a data table and produce a data frame in R.

`read.csv(file, sep=" ",head=F)`

read.xport: Read a SAS XPORT Format Library

read.dbf: Read a DBF File

write.dbf: Write a DBF File

write.table: To output an R data frame or an R matrix into a text file data table

Example:

```
pima = read.table("PIMA.txt",sep=",",head=T)
write.table(pima,file="c:\\Documents and Settings\\al\\My Documents\\587\\PIMA1.txt",row.names=F)
```

save and load:

```
save(x,y,z, file="C:/saved.txt", compress=F)
save.image(file="C:/ .RData", compress=F)
load( file="C:/saved.txt" )
dump()
source()
```

MORE DATA MANIPULATION

Sorting:

```
sort.list(pima$PEDI)
pima[sort.list(pima$PEDI), ] [1:20, ]
```

Transformations:

```
par(mfrow=c(3,3))
apply(pima[,-9],2,hist)
lPRG = log(pima$PRG)
lPRG = log(1+pima$PRG)
hist(lPRG)
pima$lPRG = lPRG
```

LOGICAL OPERATORS: `>` , `>=` , `<` , `<=` , `==` , `!=` , `&` , `|` or `! Not`

```
x = rnorm(10)
u = x > 0
u
```

```
x[u]
x
```

CONDITIONAL

```
x <- rnorm(10)
u <- sum(x)
if( u < 0) { x <- -x }
```

LOOPS

```
for loop:
z <- matrix(rnorm(200),20,10)
mean.samp <- NULL
for(i in 1:10) {
mean.samp[i] <- mean(z[,i])
}
stem(mean.samp)
while loop:
while(condition is true) { do this }
repeat { do this ; if (this) break }
apply( x, 1, fun)
sapply( x, fun)
tapply( x, y, fun)
It is very important to vectorize the code.
```

OBJECT ORIENTED PROGRAMMING

Classes and Methods

```
> print
function (x, ...)
UseMethod("print")
<environment: namespace:base>
> methods(print)
[1] print.act*
[2] print.anova
[3] print.aov*
[4] print.aovlist*
[5] print.ar*
[6] print.Arima*
[7] print.arima0*
[8] print.AsIs
[9] print.Bibtex*
[10] print.by
[11] print.check_code_usage_in_package*
[12] print.check_demo_index*
[13] print.check_make_vars*
> class(x)
```

FUNCTIONS

```
fourmom <- function(x) {
m1 <- c(mean(x))
m2 <- mean(x^2)
m3 <- mean(x^3)
m4 <- mean(x^4)
list(m1=m1,m2=m2,m3=m3,m4=m4)
```

```

}
x <- rnorm(10)
fourmom(x)
```

Homework:

Write your own function that takes the median of the rows of a matrix without using loops or apply.

Exercise:

Take the following function. Can you tell me what it does?

```
cmean <- function(x) {
  n <- nrow(x)
  p <- ncol(x)
  nax = is.na(x)
  nna <- rep(1,n) %*% nax
  i <- nna > 0
  result = rep(NA,p)
  result[!i] <- rep(1,n) %*% x[,!i] / n
  result
}
```

Modify the function so it will allow you to remove missing values.

Answer:

```
cmean <- function(x, na.rm=T) {
  n <- nrow(x)
  if( na.rm) {
    nax = is.na(x)
    nna <- rep(1,n) %*% (!nax)
    x[nax] <- 0
    return( c(rep(1,n) %*% x / nna))
  }
  else return(c( rep(1,n) %*% x /n))
}
```

PLOTS:

High level:

barplot() to draw bar-plots

```

data(VADeaths, package = "base")
barplot(VADeaths, plot = FALSE)
barplot(VADeaths, plot = FALSE, beside = TRUE)
mp <- barplot(VADeaths) # default
tot <- colMeans(VADeaths)
text(mp, tot + 3, format(tot), xpd = TRUE, col = "blue")
#
barplot(VADeaths, beside = TRUE,
col = c("lightblue", "mistyrose", "lightcyan",
"lavender", "cornsilk"),
legend = rownames(VADeaths), ylim = c(0, 100))
title(main = "Death Rates in Virginia", font.main = 4)
#
hh <- t(VADeaths) [, 5:1]
mybarcol <- "gray20"
```

```

mp <- barplot(hh, beside = TRUE,
col = c("lightblue", "mistyrose",
"lightcyan", "lavender"),
legend = colnames(VADeaths), ylim= c(0,100),
main = "Death Rates in Virginia", font.main = 4,
sub = "Faked upper 2*sigma error bars", col.sub = mybarcol,
cex.names = 1.5)
segments(mp, hh, mp, hh + 2*sqrt(1000*hh/100), col = mybarcol, lwd = 1.5)
stopifnot(dim(mp) == dim(hh))# corresponding matrices
mtext(side = 1, at = colMeans(mp), line = -2,
text = paste("Mean", formatC(colMeans(hh))), col = "red")
plot() to draw scatter-plots
x <- rnorm(30)
y <- rcauchy(30)
abline(0,1)
plot(x,y, pch=16, cex=0.7, xlab="from Normal",ylab="from
Cauchy",main="Random Data")
abline(0,1)

```

LOW LEVEL COMMANDS

```

par(mfrow=c(2,2))
mm <- array(c(1, 2), dim = 2)
nf <- layout(mm, 4, c(5, 3), TRUE)
plot(x,y)
hist(x)
par(fig=0,0.5,0,1)
par( mar=c(2,2,1,1))

```

Pairwise scatter plots:

pairs(state.x77)

3D Plots: Are sometimes useful but may need animation

Conditional plots

```

(In R) data(state)
attach(data.frame(state.x77))#> don't need `data' arg. below
coplot(Life.Exp ~ Income | Illiteracy * state.region, number = 3,
panel = function(x, y, ...) panel.smooth(x, y, span = .8, ...))
detach() # data.frame(state.x77)

```

Parallel Plot: Graph of a multivariate dataset where the observations are represented by lines.

Objectives:

1. To visualize comparisons between multivariate data groups.
2. Help assess the quality of classification tools
3. To find data clusters and outliers.

parallel(~ state.x77 | state.region)

Using the Crime dataset:parallel(~X[,1:4])

```

hist.inv <- function(x,side=1) {
gg <- hist(x,plot=F)
con <- max(gg$counts)*1.05
if( side==1) {
plot(range(gg$breaks),c(0,con),type="n", axes=F,xlab="",ylab="")

```

```

axis(2,con -( i <- pretty(gg$counts)),i)
for(i in 1:length(gg$counts)) rect(gg$breaks[i],con-
gg$counts[i],gg$breaks[i+1],con)
}
if(side==2) {
plot(c(0,con),range(gg$breaks),type="n", axes=F,xlab="",ylab="")
axis(1,con -( i <- pretty(gg$counts)),i)
for(i in 1:length(gg$counts)) rect(con-
gg$counts[i],gg$breaks[i],con,gg$breaks[i+1])
}
}

superplot <- function(x,y) {
par(mar=c(1.1,1.1,1,1))
nf <- layout(array(c(2,4,1,3), dim=c(2,2)), c(1,3), c(3, 1), TRUE)
plot(x,y)
hist.inv(y,2)
hist.inv(x) ; frame()
}
x = pima[,2]; y= pima[,3]; superplot(x,y)

##Another way of doing it

superplot2 = function(x,y) {
xhist <- hist(x, plot=FALSE)
yhist <- hist(y, plot=FALSE)
top <- max(c(xhist$counts, yhist$counts))
xrange <- range(x)
yrange <- range(y)
nf <- layout(matrix(c(2,1,0,3),2,2,byrow=TRUE), c(1,3), c(3,1), TRUE)
layout.show(nf)
par(mar=c(1,1,1,1))
plot(x, y, xlim=xrange, ylim=yrange, xlab="", ylab="")
par(mar=c(1,1,1,1))
barplot(yhist$counts, xlim=c(top, 0), space=0, horiz=TRUE) # !
par(mar=c(1,1,1,1))
barplot(xhist$counts, ylim=c(top,0), space=0) #!ylim!
invisible()
}

x <- pmin(3, pmax(-3, rnorm(50)))
y <- pmin(3, pmax(-3, rnorm(50)))

```

LIBRARIES: mva- Multivariate Analysis. Principal Components

```

library()
library(cluster)
## the variances of the variables in the
## USArests data vary by orders of magnitude
data(USArrests)
(pc.cr <- princomp(USArrests))
princomp(USArrests, cor = TRUE)
princomp(scale(USArrests, scale = TRUE, center = TRUE), cor = FALSE)
summary(pc.cr <- princomp(USArrests))
loadings(pc.cr)

```

```
plot(pc.cr) # does a screeplot.
biplot(pc.cr)
```

STATISTICAL MODELS: ANOVA

Annette Dobson (1990) "An Introduction to Generalized Linear Models".

Page 9: Plant Weight Data.

```
ctl <- c(4.17,5.58,5.18,6.11,4.50,4.61,5.17,4.53,5.33,5.14)
trt <- c(4.81,4.17,4.41,3.59,5.87,3.83,6.03,4.89,4.32,4.69)
group <- gl(2,10,20, labels=c("Ctl","Trt"))
weight <- c(ctl, trt)
anova(lm.D9 <- lm(weight ~ group))
summary(lm.D90 <- lm(weight ~ group - 1))# omitting intercept
summary(resid(lm.D9) - resid(lm.D90)) #- residuals almost identical
opar <- par(mfrow = c(2,2), oma = c(0, 0, 1.1, 0))
plot(lm.D9, las = 1) # Residuals, Fitted, ...
par(opar)
```

Smoothing Splines

```
library(modreg)
data(cars)
attach(cars)
plot(speed, dist, main = "data(cars) & smoothing splines")
cars.spl <- smooth.spline(speed, dist)
(cars.spl)
all(cars.spl $ w == table(speed)) # TRUE (weights = multiplicities)
lines(cars.spl, col = "blue")
lines(smooth.spline(speed, dist, df=10), lty=2, col = "red")
legend(5,120,c(paste("default [C.V.] => df =",round(cars.spl$df,1)),
"s( *, df = 10)'), col = c("blue","red"), lty = 1:2,
bg='bisque')
detach()
```

Writing R – packages

1. Create a package skeleton in R, using the command

```
package.skeleton(name="roots",list=c('superplot','superplot2','hist.inv')).  
Then edit the DESCRIPTION file and .Rd files appropriately
```

2. Download and install the following software:

c:\PERL: <http://www.maths.bris.ac.uk/~maman/computerstuff/Rhelp/ActivePerl-5.8.3.809-MSWin32-x86.msi>

c:\CYGWIN: <http://www.maths.bris.ac.uk/~maman/computerstuff/Rhelp/cygwin.zip>

c:\MINGWIN: <http://www.maths.bris.ac.uk/~maman/computerstuff/Rhelp/MinGW-1.1.tar.gz>

c:\program files\R\R...\\bin\HHC.EXE: <http://www.maths.bris.ac.uk/~maman/computerstuff/Rhelp/hhc.exe>

HTML HELP WORKSHOP: <http://go.microsoft.com/fwlink/?linkid=141188>

3. CHANGE PATH VARIABLE: (*Start->Settings->*)*Control Panel->System->Advanced*. Click on the *Environment Variables* button, which should be in the middle. In the lower list of variables, there should be one called *Path*. Press edit (on the lower set of buttons) and add the locations of the three new programs. There is a bit of confusion as to the whether the order in which they come matters, but we suggest

C:\Perl\bin\;C:\cygwin;[other stuff - DO NOT REMOVE];C:\mingwin\bin
NO BLANK SPACES anywhere in the path.

4. OPEN cmd window. Then go to the R folder and bin subfolder
cd C:\Program Files\R\R-2.6.1\bin
5. RUN the command to create the R-package.

Rcmd build --force --binary "*packagelocation*"

If there are any error messages that you cant resolve please bring them to class and we will deal with them.

Homework 3:

1. Fix *superplot* and *superplot2*. There is a problem with the scales on the side. The histogram scale does not exactly correspond to the scales of the axes of the scatter plot. There is a small displacement please try to fix this is you can but only after you succeed with part 2.
2. Write an R-package with these functions. *superplot*, *superplot2*, **hist.inv**.