Infection Control Related to Laboratory Safety

Dr. Kitty Fung Certificate Course in Medical Microbiology 2015, CUHK

Classification of risk groups

	Group 1	Group 2	Group 3	Group 4
Pathogenicity	Low	Moderate	High	High
infectiousness	Low	Low	Low	High
Treatment and prevention	A/V	A/V	A/V	Not A/V
Examples		Most agents in diagnostic laboratory	MTB, SARS CoV, HPAI	Viral haemorrhagic fever, smallpox

Classification of biosafety level

BSL	Agents			
1	Not known to consistently cause disease in healthy adults			
2	Associated with human disease, hazard from percutaneous injury, ingestion, mucous membrane exposure			
3	Indigenous or exotic agents with potential aerosol transmission; may have serious or lethal consequences			
4	Dangerous/exotic agents with high risk of life-threatening disease, aerosol-transmitted infections; or related agents with unknown risk of transmission			

Risk groups, biosafety levels, practices & equipment

BSL	Laboratory type	Laboratory practices	Safety equipment
1	Basic teaching, research	Good microbiological techniques	None Open bench work
2	Primary health services; diagnostic services, research	Good microbiological techniques, protective clothing, biohazard sign	Open bench PLUS biological safety cabinet for potential aerosols
3	Special diagnostic services, research	As BSL 2 PLUS special clothing, controlled access, directional airflow	Biological safety cabinet and/or other primary devices for all activities
4	Dangerous pathogen units	As BSL 3 PLUS airlock entry, shower exit, special waste	Class III biological safety cabinet, positive pressure suits, double ended autoclave (through the wall), filtered air



接觸血液、體液、分泌物 排泄物、黏膜或傷口 必須戴上手套 Wear Gloves when handling blood, body fluids, secretions, excretions, mucous membrane or non-intact skin

若有可能接觸濺出 血液或體液 必須戴上 口罩、眼罩 及穿上保護衣

Wear a Mask, Protective Eyewear and a Gown to protect yourself from splashed blood or body fluids

切勿套回 已使用的針咀

小心處理 針咀及利器

接觸血液、體液 分泌物、排泄物 黏膜、傷口, 或除下手套後 應立即潔手 **No Recapping**

Handle Sharps Carefully

Perform Hand Hygiene Immediately

after taking off gloves or handling blood, body fluids, secretions, excretions, mucous



Standard lab practices

- Restrict access to lab
- No eating, drinking, smoking, application of cosmetics & handling contact lenses in work areas
- No sandals or open-toed footwear in lab work areas
- Long hair must be tied back off the face
- Hand washing
- Decontaminate work surface & equipment after use
- Use mechanical pipetting devices
- Minimize splashes and aerosols
- Proper decontaminate of lab wastes

Protective clothing in lab area (1)

- Lab gown (solid front or wrap around gowns with cuffed sleeves) should be worn for performing lab activities
- Other PPE are worn according to risk of aerosol generation and exposure when performing specific manipulation
- PPEs should not be worn in non-laboratory areas
- Remove PPE and wash hands after use

Protective clothing in lab area (2)

Disposable gloves

- For procedures with direct or accidental contact with blood, body fluids and other potentially infectious materials
- No gloves for non-lab tasks, e.g. computer data entry, using phones, photocopying machine, fax machine

Eye/face protection

- E.g. visors, surgical mask, face shields, goggles
- For activities likely to generate splashes or sprays

Surgical masks/ N95 Respirators

- Choice of mask and respirator depends on the type of hazard
- Wear surgical mask if droplet exposure is likely
- N95 respirators
 - For aerosol generating procedures e.g. cleaning up spillage of infectious materials
 - Should choose the right size and perform fit check before each use

Use of surgical mask

















Use of N95 respirator















Donning and removal of PPEs



Removal of PPEs

- Perform at designated areas
- Do not gown down together in close proximity to another person
- Avoid touching the outer surface
- Wash hands after removal of PPEs
- Items should be properly disinfected before reuse



Hand washing

- After handling infectious materials, when contaminated, after removal of gloves, before leaving laboratory
- How?
 - Wet hands with water, apply antimicrobial soap, rub hands together for at least 20 seconds
 - Rinse and dry with paper towel

How to wash your hands??





2

Wet hands with water;

Apply enough soap to cover all hand surfaces;

1

Rub hands palm to palm;



Right palm over left dorsum with interlaced fingers and vice versa;



Palm to palm with fingers interlaced;



Backs of fingers to opposing palms with fingers interlocked;



Rotational rubbing of left thumb clasped in right palm and vice versa;



Dry hands thoroughly with a single use towel;



Rotational rubbing, backwards and forwards with clasped fingers of right hand in left palm and vice versa;



Use towel to turn off faucet;



Rinse hands with water;



Your hands are now safe.



Patient Safety

A World Alliance for Safer Health Care

SAVE LIVES Clean Your Hands

Easily missed areas during hand hygiene

Risks of transmission after exposure to certain bloodborne pathogens

- Hep B (needle stick / cut)
 6
- Hep C (needle stick / cut)
- HIV (needle stick / cut)
- HIV (mucous membrane)

- 6 30 %
- 1.8 %
- 0.3 %
- 0.09 %

Sharps precautions

DON'T touch broken glass with hands



DON'T use plastic ware



Sharps precautions

DON'T

Break, bend, recap or reuse syringes or needles

DO Use sharps containers





Seal the sharps box when ³/₄ full. Place in red bag and tag with "Biohazard labeling"





Post-exposure management

- Rinse wound or exposed skin/ mucous membrane with copious water
- Wound dressing
- Report to in-charge personnel
- Seek advice from AED



Aerosols produced by blowing out the last drop in a pipette



Minimize aerosol generation

Use of pipettes

- No mouth pipetting
- Avoid mixing by alternate suction and expulsion
- Mark-to-mark pipettes preferable to avoid expulsion of the last drop
- Submerge contaminated pipettes completely overnight in disinfectant before disposal
- Consider pipettes with cotton plugs or aerosol-barrier pipette as appropriate

Minimize aerosol generation

Centrifuge

Use sealable buckets

Open buckets after aerosols have settled (30min) or in safety cabinet

- Tissue grinders, homogenizer
- Opening ampoules of freeze-dried material
 - Wrap the ampoule in alcohol-soaked cotton wool before breaking, in safety cabinet



Class I BSC

- "Fume hood + HEPAfiltered exhaust"
- Protect staff, not specimens

----- Clean air ----- Contaminated air



Class II BSC

- Unidirectional downward flow of filtered air
- Protect staff & specimens
- Use for work with infectious agents involving:
 - Aerosols and splashes
 - Large volumes
 - High concentrations

Class II Biological Safety Cabinets



_ayout & technique



Use of Biological safety cabinets

- Minimal apparatus and materials
- Work carried out in the middle or rear part of the working surface, not on front grill
- Place discard jar within cabinet
- Avoid disruptions of laminar air flow
 - No Bunsen frame inside
 - distort air flow, damage filters
 - use disposable transfer loops or micro-incinerator
- Decontaminate materials before removal from cabinet

Use of biological safety cabinets

Position of operator

- See through the viewing screen, not under it
- Reach all area without contortions
- Minimise traffic behind the operator
- Check airflow anemometer / built-in device
- Allow to run for 10-15 min before shut off
- Regular service and maintenance

Decontamination of BSC

- Wipe surface with appropriate disinfectant after work and at the end of the day
- Avoid metal corrosive agents, e.g. chlorine-based materials

Fumigation

- At regular interval before maintenance, filter changing/testing
- After major contamination

醫療廢物 Clinical Waste □利器盒 Sharps boxes □滴血及凝有血塊的敷料

Dressing and other waste dribbling caked with blood

- 未經滅菌化驗室病菌的培養基 Unsterilized cultures & stocks from laboratory
- □具傳染性物料,如伊波拉病毒、 沙士病毒

Infectious materials, eg. Ebola virus, SARS

人體和動物組織及器官胎盤、 斷肢及動物屍體 Human and Animal Tissues, Organs and body parts

一般廢物 Municipal waste

醫療廢物分類及處置 Segregation and Disposal of Clinical Waste



Sharp boxes (containing used or contaminated sharps including syringes) 未經調道的 利閒收集箱 (內爾 化驗室病面 緩使用或受污染利 總費基或聲 當包括針筒) 用均要基 Infectious Materials e.g. Ebola virus, Severe Acute Respiratory Syndrome Coronavirus, and other contaminated materials assessed to be of significant infectious risk by health care personnel 具備染性培育,如其伊波拉病素,新重動性呼吸 系统综合症況就成每時期的間等,及其他感情器 人員經常為其相當處染性環境的影響。 Dressing and other waste dribbing and caked with blood 第四及凝有血 流的耐耗及其 也原始



 Human and animal tissues, organs and body parts 人體和動物組織、器官及身體部分



Sterilized cultures or stock from laboratory (Autoclaved) 已滅菌的化驗室病菌培 養基或儲用培養基 (高 溫滅菌) Paper hand towels 抹手紙 Wrappers

包裝紙

Nappies / incontinence pads 尿片

● Urine bags / stomal bags 尿袋及造口袋

Laboratory waste disposal

- Proper segregation
- Package
 - Bags are sealed when ³/₄ full (Don't use staples)
 - Properly labeled
- Storage
 - No accumulation in corridors, other places accessible to public (or pests and rodents)

Transport

 Dedicated bins which are cleaned regularly and after spillage

Proper packing of clinical wastes

When clinical waste bags are filled to the warning line, the "Swan-neck" method of sealing should be used before transfer to collection point

- Never use staple or unprotected metallic wire tie for sealing or tagging
- May cause injury to waste handlers or damage to bags



Clinical Waste Tag

- Every clinical waste container must bear the label "Clinical Waste"
- Labels should be securely attached to the containers, and info in the clinical waste tag clearly marked using BLACK indelible ink with information showing the origin of the waste:



- 醫院名稱
- 部門/病室/診所名稱 Dept / Ward / Clinic
- -包裝日期

Packaging Date

Name of hospital

Effluent from analytical equipment

- **1**. Trapped in bottles containing hypochlorite
- 2. Discharged directly into the waste plumbing system
 - A discharge tube shall project at least 25 cm into the pipe-work to avoid splashing
 - Water flow down the waste-pipe while the machine is operating
 - Waste system treated with 250 mL of hypochlorite (0.25% available chlorine) when the work is finished

Specimen transport (Triple packaging system)

- Primary (1°) container
 - should be properly capped, watertight and leak proof
- Secondary (2°) container
 - Request form in the pocket of the zip-lock bag
- Outer (3°) container
 - Regularly cleansed and disinfected
 - Biohazard warning label on the outside


Intra-hospital specimen transport (routine specimens only)



Contents of biological spill kit



Contents of biological spill kit

- Water repellent solid front gown
- N95 mask: 3M 1860S, 3M 1860, 3M 1862, 3M 8210
- Surgical mask
- Shoe cover
- Cap
- Face shield
- Latex gloves

- Warning sign
- Warning tape
- Virusolve+ soln (40ml) & DW (760 ml) to prepare 5% soln
- Absorbent pads & Gauze
- Forceps
- Cards
- Garbage bag (black)
- Autoclave bag

Activity spectrum of select detergents and disinfectants

+++ ++++ 0 +++ ++++	++ ++ 0 NP 0	0 + 0 NP 0	+ ++++ + 0 +	+ ++ + NP +	0 0 0 0 0	
+ 0 ++	0 NP	0 NP	+ 0	+ NP	0	
0++	NP	NP	0	NP	0	
++					-	
	0	0	+	+	0	
	++	++	++	++	+ ^(a)	
+++	++	++	++	++	0	
++	0	0	+	0 ou +	0	
Variable activity depending on components ^(b)						
+	0	0	+	0	0	
	++ Varia +	++ 0 Variable activi + 0	++00Variable activity dependir+00	++00+Variable activity depending on com+00+	++ 0 0 + 0 ou + Variable activity depending on components ^(I)	

Source: WHO

Virusolve+ : spectrum of activity

Bacteria

- Acinetobacter species
- Bacillus species
- Campylobacter jejuni
- Clostridium difficile
- Escherichia coli
- Legionella pneumophila
- Salmonella typhimurium
- Staphylococcus aurues
- Vibrio cholarae
- Yersinia enterocolitica
- Mycobacterium
 - M. avium
 - M. tuberculosis

Viruses

- Avian flu (H5N1)
- Hepatitis B and C
- HIV-1
- Polio virus
- Influenza virus
- Vaccinia virus
- Spores
 - Bacillus cereus, B. subtilis
 - Clostridium difficile
- Fungi
 - Aspergillus species
 - Candida albicans

Virusolve+ solution

- Non-flammable, Non-irritant
- Potential skin and eye irritant
- Not corrosive to metal
- A combination disinfectant & cationic surfactant
- Contains alkyl triamine to disrupt membrane and cause cell lysis
- Store upright in a cool, dry, well-ventilated area. Avoid exposure to direct sunlight or sources of heat



Handling of minor spillage

- For Group 1 and 2 pathogens (e.g. dropped swabs, drops of culture broths, slight spills of specimens)
- Pick up broken pieces of glass/sharps using forceps with gloved hands, dispose into sharps box
- Wipe spill with disposable absorbent materials wetted with freshly prepared 1% hypochlorite (e.g. 1 in 5 bleach) or 5% Virusolve+ solution, rinse with water after 1 min
- Discard used gloves and soaked cloth or paper towels as contaminated materials

Handling of spillage during centrifugation

- Switch off the machine
- For non-sealable buckets
 - Close the lid for 30 min to allow aerosols to settle before opening
 - Use forceps to pick up glass debris
- For sealable buckets
 - Open the centrifuge after it has stopped
 - Remove the buckets / contents to a BSC
- Immerse all broken tubes, glass fragments, buckets, trunnions, rotor in 5% Virusolve+ solution for 15 minutes, rinse with water
- Swab the bowl with 5% Virusolve+ solution, swab dry

Handling of major spillage

(e.g. spillage and/or breakage of laboratory cultures)

Persons involved

- Inform other workers for temporary evacuation
- Remove contaminated PPE
- Leave the room, close door and put a warning sign on the door
- Report to safety officer/ supervisor
- Self-decontamination: wash, flush
- Seek advice from ICT as appropriate

Decontamination of major spillage

- Carried out ASAP (except in negative pressure room) by trained personnel
- Put on PPE : water-resistant gown, gloves, N95 respirator, face shield
- Cover contaminated area with lint clothes, flood them with 5% Virusolve+ solution (1 in 20) for 15 min
- Wipe clean and dry surfaces with paper towel
- Gather materials into waste bags and autoclave ASAP

Handling of major spillage inside BSC

- Allow the cabinet to continue operation to contain and exhaust the aerosols
- Do not remove items from the cabinet
- Cover contaminated area with diposable absorbent materials, flood them with 5% Virusolve+ solution (1 in 20) for 15 min, swab with water and air dry. Avoid using metal corrosive disinfectants like hypochlorite.
- If the cabinet incorporates a catch basin beneath the work surface, this should be flooded with the disinfectant.
- Disinfect or autoclave contaminated items as appropriate
- Fumigate the cabinet before activity is resumed.

Handling of specimens suspected of highly pathogenic micro-organisms

Activities performed in BSL-2 facilities with BSL-2 practice

- Routine diagnostic testing of serum, blood and urine specimens
- Routine staining and microscopic analysis of fixed smears
- Routine examination of mycotic and bacterial cultures
- Pathological examination and processing of formalin-fixed / inactivated tissue
- EM studies with glutaraldehyde-fixed grids
- Molecular analysis of extracted nucleic acid preparations

Activities performed in BSL-2 facilities requiring BSL-3 practices

- Aerosols or splash generating procedures
 - e.g. sonication, vortexing, grinding or blending
- Aliquot and/or dilute specimens
- Manipulation of untreated specimens
- Inoculation of bacterial or mycological culture media
- Preparation of chemical- or heat-fixing of smears for microscopic analysis
- Nucleic acid extraction procedures involving untreated specimens

BSL-3 practices

- Conduct procedures within BSC
- Wear appropriate PPE
 - Disposable gloves, solid front or wrap around gowns with cuffed sleeves, and a surgical mask or full-face shield
- Centrifugation in sealed centrifuge rotors/ cups and unloaded inside BSC
- Decontaminate work surfaces and equipment after use e.g. diluted bleach (1 in 50)

Smear preparation

Fresh respiratory specimens for cytological examination:

- Prepare smears inside the BSC
- Fix in 95% ethanol for 30 minutes + fix in 70% ethanol or isopropyl alcohol for 15 minutes (for disinfection) or
- Fix in equal volume of 10% formalin for 2 hours before smear preparation and staining

Peripheral blood

 Smears can be incubated on a hot plate heated up to 56°C for 15-30 minutes before methanol fixation and Romanowsky staining.

Enhanced PPE

- Use when the procedure cannot be conducted within a BSC
- Includes the followings as appropriate
 - Disposable gloves
 - Solid front or wrap around gowns with cuffed sleeves
 - N95 respirator
 - Full face shield
 - Head covering , shoe cover and other physical containment devices

BSL-3 Laboratory

For diagnostic tests that involve

- Cell culture for virus isolation
- Initial characterization of viral agents recovered in cultures (e.g. specimens for avian influenza)

	BSL-2 standard + BSL-2 Practice	BSL-2 standard + BSL-3 Practice	BSL-3 standard + BSL-3 Practice			
Requirement	Open Bench	BSC	BSC			
PPE	Gloves, Gown, Surgical Mask	Gloves, Gown, Surgical Mask	Following individual laboratory's guideline			
Enhanced PPE For exceptional BSL-3 practice	N/A	Gloves, Gown, Goggles, N95 respirator, Head covering and Dedicated shoes/Shoes cover as appropriate	N/A			

Immunisation for lab staff

- Hepatitis B vaccine
- Influenza vaccine
- MMR vaccine
- Tetanus vaccine

